

Available online at www.sciencedirect.com



Tetrahedron Letters

Tetrahedron Letters 48 (2007) 8230-8233

A simple method for the alkaline hydrolysis of esters

Vassiliki Theodorou,^{a,*} Konstantinos Skobridis,^a Andreas G. Tzakos^b and Valentine Ragoussis^c

^aDepartment of Chemistry, University of Ioannina, GR-451 10 Ioannina, Greece ^bMRC, Laboratory of Molecular Biology, Hills Road, CB2 0QH Cambridge, United Kingdom ^cDepartment of Chemistry, University of Athens, Panepistimiopolis, Zografou, GR-157 71 Athens, Greece

> Received 29 June 2007; revised 3 September 2007; accepted 12 September 2007 Available online 18 September 2007

Abstract—A very mild and rapid procedure for the efficient alkaline hydrolysis of esters in non-aqueous conditions has been developed, by the use of dichloromethane/methanol (9:1) as solvent. This method conveniently provides both carboxylic acids and alcohols from the corresponding esters and sodium hydroxide in a few minutes at room temperature. A plausible reaction mechanism is proposed.

© 2007 Elsevier Ltd. All rights reserved.

Synthesis and structure determination, two of the major concerns of the organic chemist, usually involve the conversion of functional groups. Among other transformations, the saponification of esters to carboxylic acids is a very common organic transformation and one of the most extensively studied reactions in chemistry.^{1,2}

Ester derivatives are widely present in many drugs, natural products and synthetic compounds. Furthermore, they are frequently used for the protection of carboxylic acids and alcohols, which, as synthons, are unmasked late in the synthesis.

Saponification processes typically involve the use of aqueous solutions, combining water and other water miscible organic solvents, such as methanol, ethanol, tetrahydrofuran, dioxane and dimethoxyethane in order to dissolve both, esters and hydroxides (KOH, NaOH, LiOH). The reaction temperature varies from room temperature to the boiling point of the mixture, while the length of the reaction time varies from 30 min to 24 h, or more. A relatively large excess of hydroxide is normally used, with a concentration chosen to be, usually, between 0.1 N and 2 N.

The rate of a saponification reaction is altered by steric and electronic effects,³ as well as by the solvent,⁴ due to specific interactions of solvent molecules with the reactants. Hydroxylic solvents interact strongly with anions and increase the energy barrier for reactions that involve the approach of a negative charge to a neutral system. A direct HO⁻/ester nucleophilic collision in an aqueous environment must involve a great amount of desolvation energy, since the hydroxide ion is strongly solvated by water molecules.⁵

Changing the traditional protic solvents of saponification reactions to other solvents that do not stabilize the reactants, seems to be a rather good modification, since in less polar aprotic solvents the reaction goes faster.

In a recent study⁶ on the selective enrichment of the β -COOH of Ac-Arg(Tos)-Gly-Asp(OBn)-Resin with oxygen-17, during solid phase peptide synthesis (SPPS), it has been established that the solvent system dichloro-methane/methanol, compatible with SPPS, in a ratio of about 6:1–9:1, was optimal for successful saponification of the side chain of the peptide with Na¹⁷OH. Methanol was used in order to dissolve sodium hydroxide. Methanolysis was, also, observed to some extent in the presence of higher-levels of methanol.

This interesting observation encouraged us to study and develop an efficient and practical saponification process

Keywords: Saponification of esters; Mechanism; Non-aqueous conditions; Methanolysis; Selective enrichment of COOH; Carboxylic acids; Alcohols.

^{*} Corresponding author. Tel.: +30 26510 98591; fax: +30 26510 98682; e-mail: vtheodor@cc.uoi.gr

^{0040-4039/\$ -} see front matter @ 2007 Elsevier Ltd. All rights reserved. doi:10.1016/j.tetlet.2007.09.074

in non-aqueous conditions that would be applied to a variety of esters (Eq. 1).

$$\begin{array}{c} \text{RCOOR'} \xrightarrow{1. \text{ NaOH, rt}} \\ (\text{CH}_2\text{Cl}_2/\text{CH}_3\text{OH} \sim 9:1) \\ \hline 2. \text{ H}_3\text{O}^+ \end{array} \right) \xrightarrow{\text{RCOOH + R'OH}} (1)$$

To the best of our knowledge, there are only a few reports⁷ on non-aqueous saponification conditions. Among the various approaches, $(CH_3)_3COK$ in DMSO, or in dry Et₂O with the presence of 1 equiv of water, has been reported for the hydrolysis of hindered esters,^{7a,b} while Ba(OH)₂·8H₂O in methanol has been used to deprotect simple alkyl esters,^{7c} under conditions suitable for parallel synthesis. Recently, sodium trimethylsilano-late (TMSONa)^{7d} has been utilized for the selective removal of methyl esters on the chain of Glu or Asp on solid support. More recently, the use of hydroxide ion-exchange resin has been investigated for ester hydroly-sis^{7e} in various organic solvents, but the proposed procedure was time consuming. A microwave-enhanced hydrolysis of esters,^{7f} using KF–Al₂O₃ under solvent free conditions has also been described.

Procedures allowing simple, efficient and reliable ester hydrolysis, without racemizations or other undesirable side reactions would be advantageous and very helpful to the broad scientific community. In order to establish a gentle, time and temperature effective methodology, without large excess of alkali or high concentrations of reactants, avoiding undesirable by-products, we sought to perform saponification experiments on several types of esters using CH_2Cl_2 as the reaction solvent, containing methanol in a ratio $CH_2Cl_2/CH_3OH \sim 9:1$.

A range of several carboxylic esters was saponified quantitatively in dichloromethane using NaOH (dissolved in MeOH) with a final normality of 0.1-0.5 N, for a short time at rt (Table 1). The aprotic solvents Et₂O, THF and dioxane demonstrated comparable efficiency, however, CH₂Cl₂ was the best choice, since the reaction was apparently faster and the work-up easier.

From the results summarized in Table 1, it is obvious that the proposed hydrolysis protocol⁸ works efficiently in all cases. Minor differentiations could be observed depending on the substrate structure. As expected, the reaction rate depends on the concentration and the ratio of the reactants, as well as on steric and electronic effects.

The described methodology provides an efficient laboratory procedure for ester hydrolysis and the possibility of

Table 1. Reaction conditions for the alkaline hydrolysis of esters to carboxylic acids

Entry	Ester	RCOOR'/NaOH	NaOH (N)	Time ^a	RCOOH ^b	
					Mp (°C)	$\left[lpha ight] _{\mathrm{D}}^{20}$
1	Ac–Pro–OMe ^c	1:1.1	0.1	50 min	116–118 (115–117) ^{10b}	−167° (−171),
		1:5	0.2	15 min		c 1 in CHCl ₃ ^{12a}
2	Boc-Tyr(2,6-di-Cl-Bzl)-OMec	1:1.1	0.1	1 h 20 min	103–104 (105) ^e	+20.2° (+20),
		1:2	0.1	45 min		c 2 in EtOH ^e
		1:4	0.1	30 min		
3	Boc-Glu(OAll)-OH	1:2	0.1	15 min		-14.1° (-14.5),
		1:3	0.1	10 min		c 1 in MeOH ^e
4	Boc-Asp(OBzl)-OH	1:3	0.1	10 min		-4.8° (-5.0),
						c 1 in MeOH ^e
5	Fmoc–Asp(OMe)–OH ^d	1:3	0.1	10 min	_	
6	CH ₂ =C(CH ₃)-COOMe	1:1.5	0.5	15 min	_	
7	CH ₃ CH=CHCOOEt	1:1.5	0.3	15 min	70–72 (70–72) ^e	
8	PhCH=CHCOOEt	1:1.5	0.2	55 min	130–132 (133) ^e	
9	PhCOOMe	1:1.5	0.2	1 h 20 min	120–122 (122–123) ^e	
10	n-C ₄ H ₉ CH=CHCH ₂ COOEt	1:2	0.2	30 min		
11	<i>n</i> -C ₅ H ₁₁ CH=CHCH ₂ COOMe	1:2	0.2	30 min	_	
12	RCH(COOEt) ₂					
	(a, R = H)	1:1.1	0.5	35 min	130–133 (134–135) ^e	
	(b, R = Me)			45 min	128–131 (129–132)	
	(c, R = n-Bu)			1 h 50 min	$100-101 (102)^{12c}$	
	(d, R = Bzl)			2 h	110–114 (117–120) ^e	
13	Methyl (3,5-dibenzyloxy)benzoate	1:8	0.5	4 h	208–209 (210-1) ^{12d}	
14	Isopropyl myristate	1:2	0.2	$\sim 2 h$	55–57 (57–58) ^{12e}	
15	Tripalmitin	1:2	0.2	$\sim 2.5 \text{ h}$	$62-64 (65)^{12f}$	
16	CH ₃ COOPh	1:1.5	0.3	5 min		
17	$CH_3COOCH_2CH=C(CH_3)_2$	1:1.5	0.2	15 min		
18	Geranyl phenylacetate	1:2	0.2	$\sim 4 h$	75–77 (77–78) ^e	
19	Benzyl pivalate	1:2	0.2	$\sim \! 10 \ h$	_	
20	2-Phenylethyl pivalate	1:2	0.2	$\sim \! 12 h$	_	

^a Monitored by TLC until the starting material was exhausted.

^b The literature melting points and specific rotations are given in parentheses.

^c The methyl esters of these amino acids were prepared from the corresponding protected amino acids with diazomethane.

^d With simultaneous Fmoc-deprotection.

^e These values have been derived from the pure products supplier (Fluka Laboratory Chemicals).



Figure 1. Suggested mechanism for the saponification of esters in CH₂Cl₂/MeOH ~9:1 (path A: direct, path B: through methanolysis).

isolating both products,^{8,9} acids (entries 1–15, 18) and/ or alcohols (entries 16–20).

The saponification of several discrete classes of natural products, such as, triglycerides, terpenoids and amino acid derivatives, received more attention and a detailed monitoring of the optimum conditions of time and reagents concentration at rt has been performed.

The hydrolysis conversion was quantitative for all the compounds studied, as shown in Table 1. The yields of recovered pure products were $\sim 80-90\%$, except for entries 1, 3 and 4, where the yields were about 70–75%, due to the high solubility of products in both water and organic solvents. For cases 5, 12a, 16 and 17 yields were not determined, as the reaction products were, also, very soluble in water and without special interest.

Importantly, the saponification of the methyl esters of orthogonally protected amino acids (entries 1 and 2) was carried out without racemization, as evidenced by their observed specific rotations $[\alpha]_{D}$. In the case of side-chain ester derivatives of amino acids (entries 3-5), one more equivalent of NaOH was required, due to the α -COOH group. It is, also, of interest that the saponification of β , γ -unsaturated esters (entries 10 and 11) was accomplished without any isomerization to the corresponding α,β -unsaturated acids, as shown by their ¹H NMR spectra, compared with those of authentic samples. The saponification of diethyl malonates^{3c} to malonic acids, was found to proceed relatively rapidly, depending on the substituent R, without any decarboxylation. On the other hand, longer reaction times, or more harsh reaction conditions were required for the sterically hindered esters, especially for entries 13 and 18-20, while the esters of fatty acids (14 and 15) reacted at a moderate rate (Table 1).

The ¹H NMR data of all the isolated products, without further purification, were identical to those of the expected pure compounds.

Conversion reactions for entries 1, 2, 9 and 12a were, also, performed with equivalent amounts of MeONa and $H_2^{17}O$, instead of NaOH, in order to produce

 $Na^{17}OH$ for the selective enrichment of the COOH group.¹⁰

The method appears to be fairly general, allowing quantitative hydrolysis of both alkyl and aryl esters. Moreover, it can be applied in cases where aqueous workup must be avoided, since the sodium salt RCOONa can be isolated pure,⁹ while the alcohol can be extracted. It is well known that triglycerides in olive oil samples,^{11a} cinnamates,^{3,11b} prolinates,^{11c} other amino acids,^{11d} malonates,^{12b,c} etc., are saponified under relatively more vigorous reaction conditions, compared with these of the present work.

A plausible mechanistic process, very similar to the widely accepted classical mechanism² and consistent with our results, can be proposed (Fig. 1). The formation of a tetrahedral intermediate would be achieved via a nucleophilic attack on the carbonyl carbon by the HO⁻ with assistance by a methanol molecule (path A). Moreover, a parallel process, where nucleophilic interaction of a methanol molecule, assisted by a hydroxide ion, could be responsible for the observed methyl ester,⁶ which is, subsequently saponified (path B). In the less polar solvent CH₂Cl₂, used in this work, the HO⁻/ester nucleophilic attack is favored, and the reaction accelerated, due to extensive desolvation of the hydroxide ions.

In summary, a simple, rapid and gentle saponification method has been established, which involves the use of CH_2Cl_2 as the reaction solvent, containing about onetenth its volume of methanol, and low concentrations of the hydroxide at room temperature. This methodology constitutes an advantageous alternative protocol, compared to the existing methods, for the alkaline hydrolysis of esters.

References and notes

 (a) Euranto, E. K. In *Chemistry of Carboxylic Acids and Esters*; Patai, S., Ed.; Interscience: New York, 1969; pp 505–588; (b) Beigley, J. R.; Hutton, A. T.; McLean, W. D. *Org. Process Res. Dev.* **1999**, *3*, 224–226; (c) Höller, U.; Wolter, D.; Hofmann, P. V.; Spitzer, P. J. Agric. Food Chem. 2003, 51, 1539–1542; (d) Berg, H.; Turner, C.; Dahlberg, L.; Mathiasson, L. J. Biochem. Biophys. Methods 2000, 43, 391–401.

- (a) Marlier, J. F. J. Am. Chem. Soc. 1993, 115, 5953–5956;
 (b) Madura, J. D.; Jorgensen, W. L. J. Am. Chem. Soc. 1986, 108, 2517–2527;
 (c) Zhan, C.-G.; Landry, D. W.; Ornstein, R. L. J. Am. Chem. Soc. 2000, 122, 2621–2627, and 7672–7676;
 (d) Mata-Segreta, J. J. Am. Chem. Soc. 2002, 124, 2259–2262.
- (a) Wang, N.; Wang, C.-H. J. Org. Chem. 1971, 36, 3178– 3179; (b) Hoffman, M. K.; Berliner, E. J. J. Org. Chem. 1970, 35, 745–749; (c) Bryant, W.; Smith, D. M. J. Am. Chem. Soc. 1936, 58, 1014–1017.
- (a) Gallagher, G. A.; Miller, J. G.; Day, A. R. J. Am. Chem. Soc. 1957, 79, 4324–4327; (b) Cramar, C.; Hawkins, C.; Truhlar, D. J. Chem. Soc. Faraday Trans. 1994, 90, 1802–1804; (c) Xie, D.; Zhou, Y.; Xu, D.; Guo, H. Org. Lett. 2005, 7, 2093–2095.
- (a) Matta, M. S.; Toenjes, A. A. J. Am. Chem. Soc. 1985, 107, 7591–7596; (b) Kovach, I. M.; Elrod, J. P.; Schowen, R. L. J. Am. Chem. Soc. 1980, 102, 7530–7534; (c) Hæffner, F.; Hu, C.-H.; Brinck, T.; Norin, T. J. Mol. Struct. (Theochem.) 1999, 459, 85–93.
- Theodorou-Kassioumis, V.; Biris, N.; Sakarellos, C.; Tsikaris, V. *Tetrahedron Lett.* 2001, 42, 7703–7705.
- (a) Chang, F. C.; Wood, N. F. *Tetrahedron Lett.* **1964**, *5*, 2969–2973;
 (b) Gassman, P. G.; Schenk, W. N. *J. Org. Chem.* **1977**, *42*, 918–920;
 (c) Anderson, M. O.; Moser, J.; Sherrill, J.; Guy, R. K. *Synlett.* **2004**, 2391–2393;
 (d) Minta, E.; Boutonnet, C.; Boutard, N.; Martinez, J.; Rolland, V. *Tetrahedron Lett.* **2005**, *46*, 1795–1797;
 (e) Morwick, T. M. *J. Comb. Chem.* **2006**, *8*, 649–651;
 (f) Kabalka, G. W.; Wang, L.; Pagni, R. M. *Green Chem.* **2001**, *3*, 261–262.
- 8. General experimental procedure for the saponification reaction: To a solution of the ester (1 equiv), in a CH₂Cl₂/CH₃OH (~9:1, v/v) mixture, was added a methanolic solution of NaOH 2–3 N (1–4 equiv), with the final concentration of the alkali being about 0.1–0.2 N. After 2–3 min of stirring, the solution became cloudy and the

sodium salt of the carboxylic acid started to precipitate. The mixture was stirred until all the ester was consumed to give a large amount of white precipitate. The reaction was monitored by TLC for the disappearance of the starting ester (Table 1). The solvents were then removed under vacuum, the residue was diluted with water and the aqueous solution was extracted with diethyl ether in order to isolate the water insoluble alcohol (16–20), or/and to remove any negligible amount of unreacted ester. The aqueous phase was then cooled, acidified to pH 2–3 with dilute HCl or NaHSO₄ and extracted with AcOEt or Et₂O or CH₂Cl₂. The combined organic layers were dried (Na₂SO₄) and the solvent was removed to afford the acid (1–15 and 18).

- 9. Alternatively, the white salt RCOONa, precipitated during the reaction, or after the addition of diethyl ether for the lipophilic products, can be isolated pure by filtration, washed with CH_2Cl_2 and conveniently manipulated.
- (a) Theodorou, V.; Troganis, A.; Gerothanassis, I. P. *Tetrahedron Lett.* 2004, 45, 2243–2245; (b) Theodorou, V., unpublished results.
- (a) Ballesferos, E.; Gallego, M.; Vercarcel, M. Anal. Chim. Acta 1993, 282, 581–588; (b) Sawada, Y.; Kayakiri, H.; Abe, Y.; Imai, K.; Mizutani, T.; Inamura, N.; Asano, M.; Aramori, I.; Hatori, C.; Katayama, A.; Oku, T.; Tanaka, H. J. Med. Chem. 2004, 47, 1617–1630; (c) Pascal, R.; Sola, R. Tetrahedron Lett. 1998, 39, 5031–5034; (d) Pasto, M.; Moyano, A.; Pericas, M.; Riera, A. J. Org. Chem. 1997, 62, 8425–8431.
- (a) Sakai, R.; Rinehart, K. L.; Kishore, V.; Kundu, B.; Faircloth, G.; Gloer, J. B.; Carney, J. R.; Namikoshi, M.; Sun, F.; Hughes, R. G., Jr.; Gravalos, D. G.; de Quesada, T. G.; Wilson, G. R.; Heid, R. M. J. Med. Chem. 1996, 39, 2819–2834; (b) Clive, D. L. J.; Hisaindee, S. J. Org. Chem. 2000, 65, 4923–4929; (c) Vogel's Textbook of Practical Organic Chemistry, 5th ed; Longman: London, 1989; (d) Hawker, C. J.; Lee, R.; Fréchet, J. M. J. J. Am. Chem. Soc. 1991, 113, 4583–4588; (e) Storck, G.; Brizzolara, A.; Landesman, A.; Szmuszkovicz, J.; Terrell, R. J. Am. Chem. Soc. 1963, 85, 207–222; (f) Shukla, Y. N.; Thakur, R. S. Phytochemistry. 1985, 24, 1091–1092.